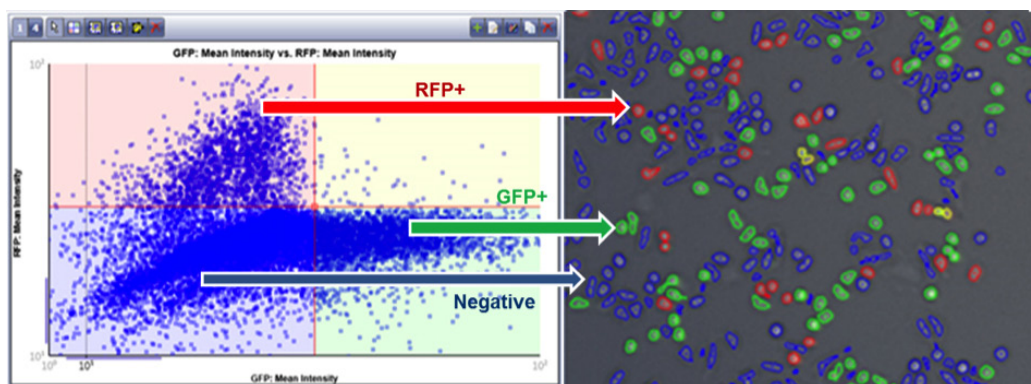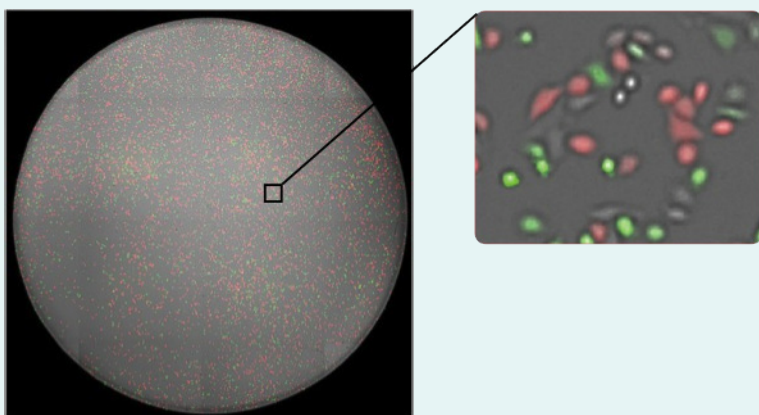# Multiplex Fluorescence Assays for Adherence Cells without Trypsinization

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The combination of a bright field and three fluorescent channels allows the Celigo to perform many multiplexed assays. A gating interface similar to flow cytometry provides great flexibility in data analysis. In addition, images of gated cells are displayed and updated with the gating operation.

- Reduce sample preparation and number of cells per test (96 or 384 wells)
- Cells stay in multi-well plates without trypsinization
- Images provide visualization of cell morphology, fluorescent distribution for assay development and quality control
- Gating is reflected on cell images

Fluorescence Proteins

Whole well view GFP/RFP transfected Hela cells in a 96-well plate.

- Identify cells in bright field image
- Measure fluorescent protein signals in green, and red channels
- Label-free, non-invasive – no need to trypinsize adherence cells
- Quantify fluorescent protein signals on a cell-by-cell basis repeatedly on the same plate providing temporal data.
- Add propidium iodide for viability of GFP transfect cells in the same well

Transfection and Transduction Optimization using Celigo Adherent Cell Cytometer

The combination of a bright field and three fluorescent channels allows the Celigo to perform many multiplexed assays. A gating interface similar to flow cytometry provides great flexibility in data analysis. In addition, images of gated cells are displayed and updated with the gating operation.

Example 1. GFP Transfection Optimization in 96 Well Plates Without Trypinization

HeLa cells were transfected with a range of concentrations of a plasmid encoding turbo-GFP1 and seeded out into a 96-well plate. To monitor cell death, propidium iodide was added to the wells in some experiments. The same plate was imaged daily over a 5-day period.
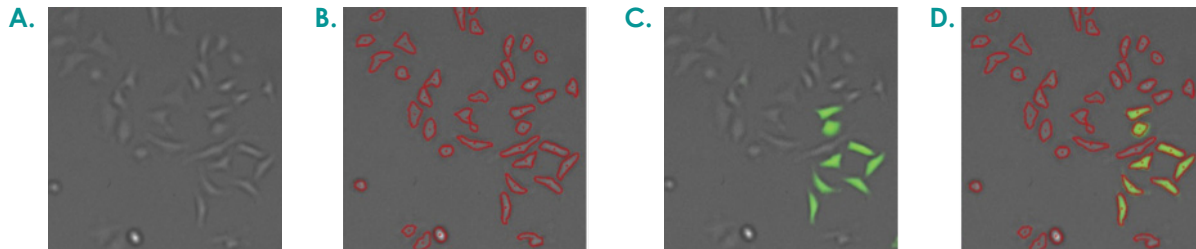

Figure 1. Cell Images of GFP transfected Hela in 96-well plate. (A) Bright field cell image including transfected and non-transfected cells. (B) Celigo image process software identifies all the cells using bright field image, indicated by the red outline. (C), (D) Within this red outline, green fluorescence intensity was measured to gate cell populations   produce transfection efficiency.
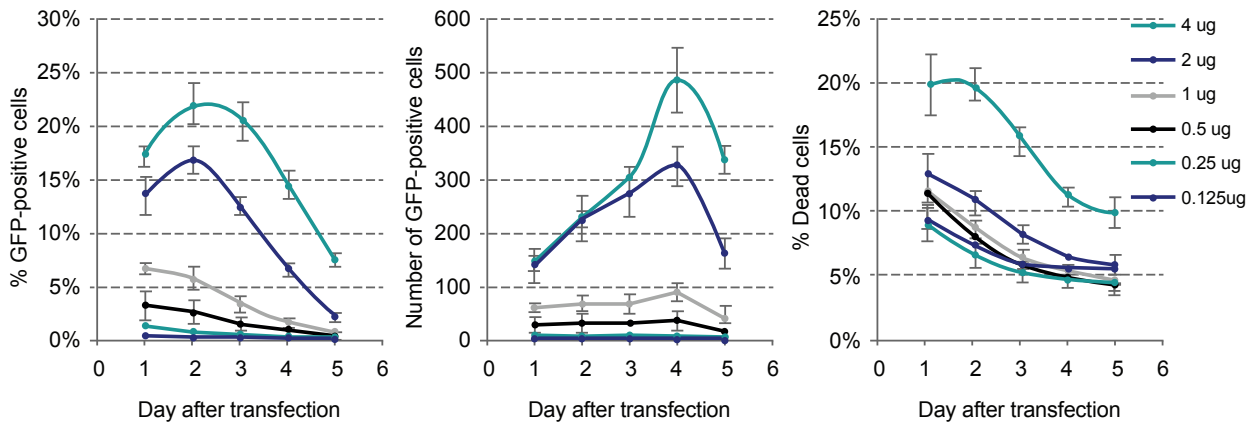

Figure 2. GFP-Positive Cell Quantification. Left: Percentage of GFP-positive cells vs. day after transfection. Middle :GFP-positive cell counts. Right: Percentage of dead cells (propidium iodide-positive). Error bars indicate standard deviation.

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Example 2. Measure % RFP Positive Cells in Samples Containing GFP, RFP and Non-Expressing Cells

96-well containing samples with varied % of RFP, GFP and non-expressing Hela cells were counted using Celigo. Bright field, green fluorescence and red fluorescence channels were used.

Step 1. Acquire & View Images on Celigo Image Cytometer

**Bright field image of well D6, zoomed view.**
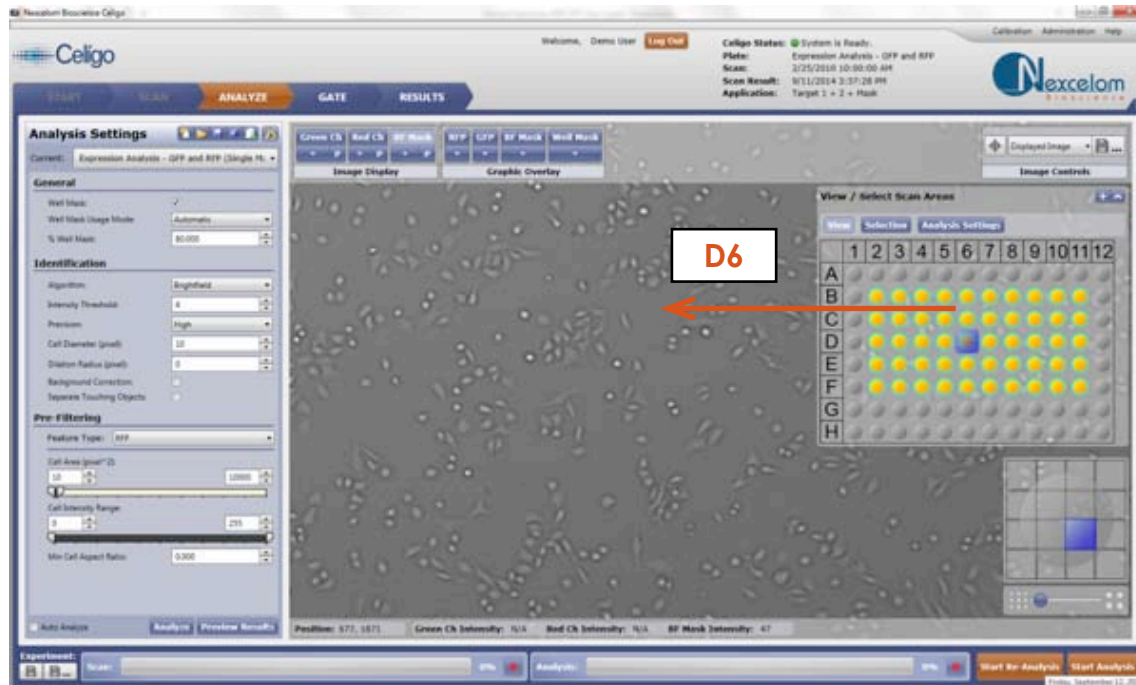
RFP fluorescence image of well D6, zoomed view.

Step 2. Gate and Circle Counted Cells Automatically

Overlay of bright field, GFP+ and RFP+ cells identified by Celigo image processing software. GFP vs RFP intensity scatter plot is also shown. When the gate on the scatter plot is modified by dragging, the counted cell image is updated automatically.
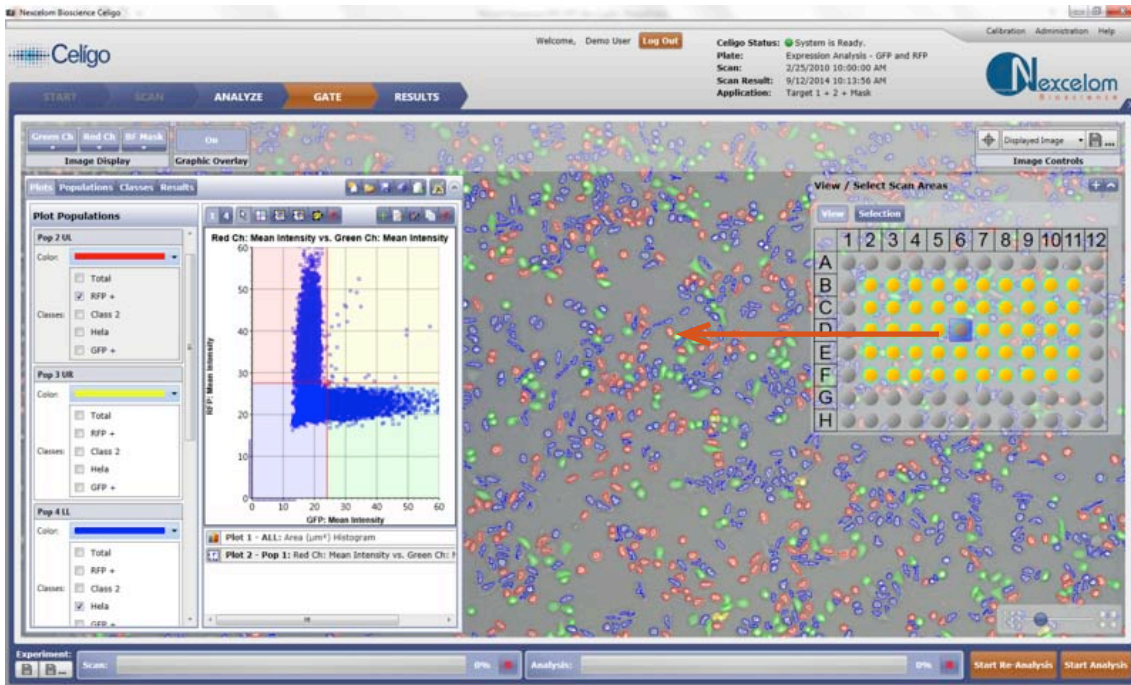
View whole well cell images
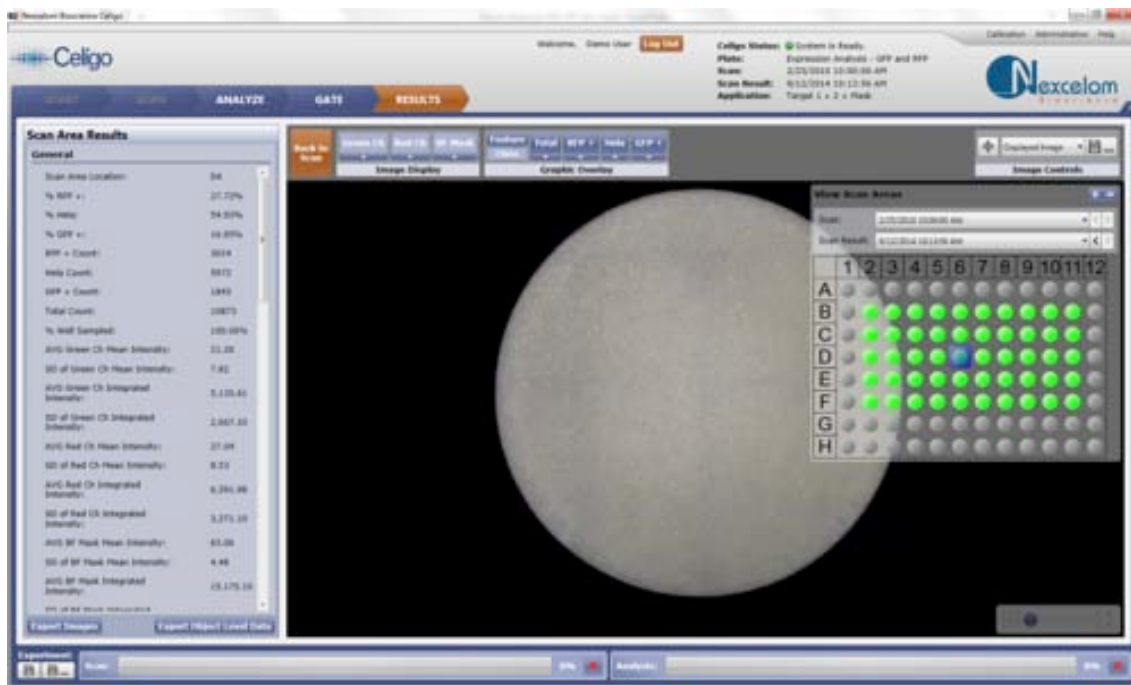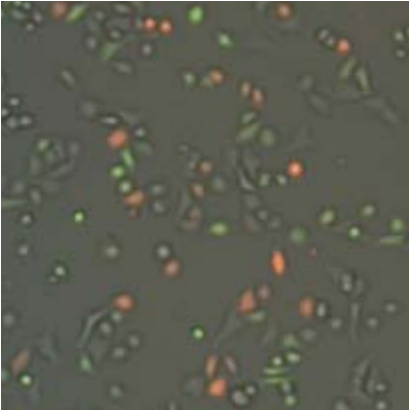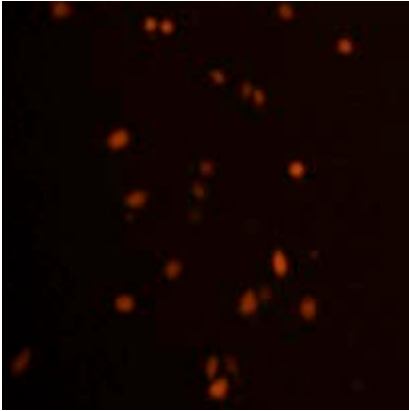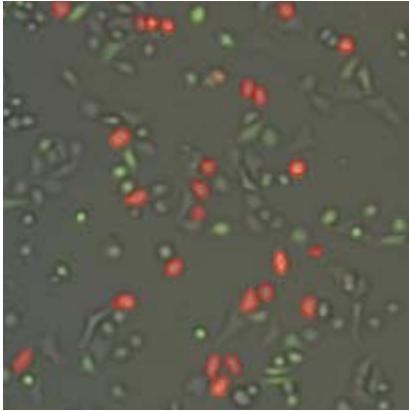Double click any well on the well map to display whole well images. Zoom in to see details as listed in Table 1.
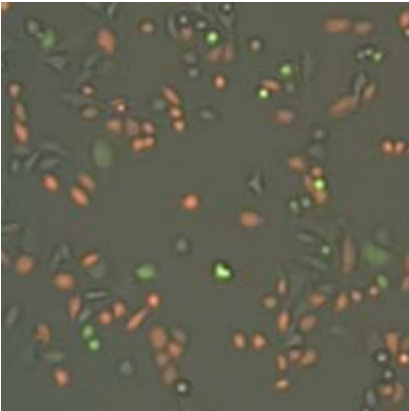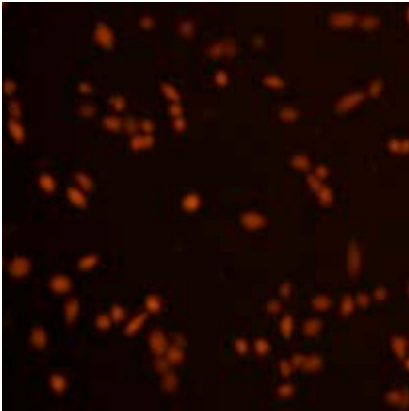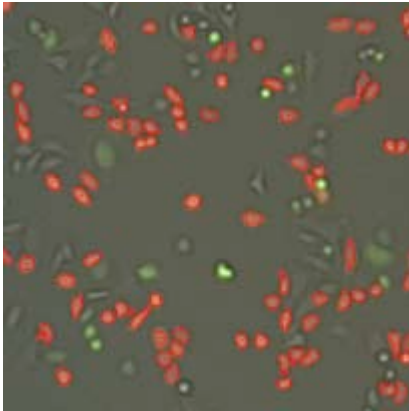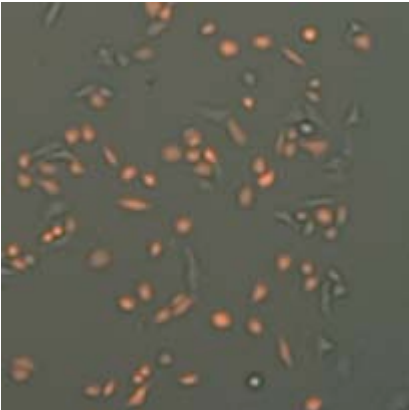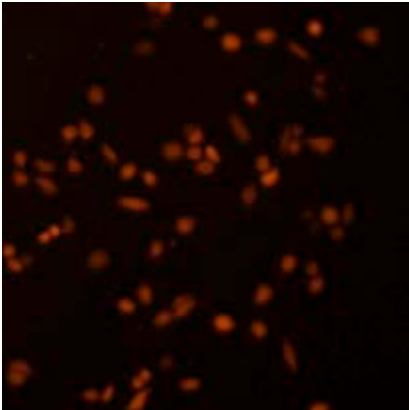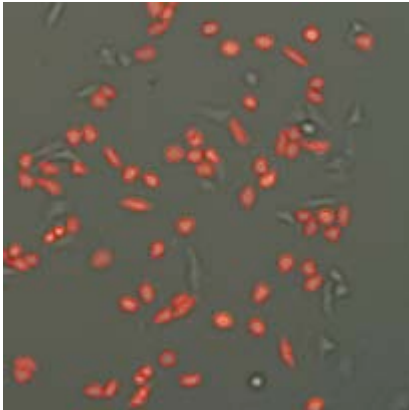
Table 1. Cell image examples from wells with varied RFP+ cell populations.

Cell Images		
GFP+, RFP+, GFP-RFP-	RFP+	Counted RFP+

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Step 3. Review Plate Level Results for Each of the Wells in the 96-Well Plate
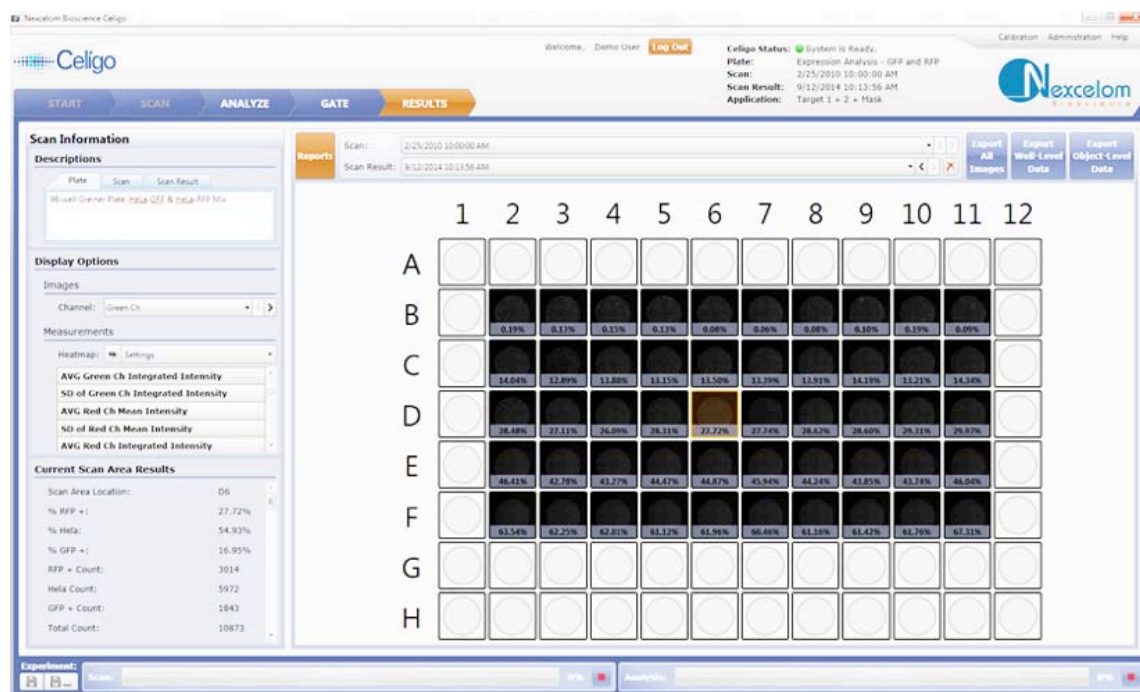
Reported Data	%	Count	Mean Intensity AVE & SD	Integrated Intensty AVE & SD
Total		•		
RFP+	•	•	•	•
GFP+	•	•	•	•
Non-FL Hela	•	•	•	•
Green Channel			•	•
Red Channel			•	•
Brightfield Mask Channel			•	•

More plots are available from Celigo software, as listed in Table 2.

Table 2. Example plots

GFP vs. RFP Scatter Plot	RFP Intensity Histogram
13% RFP	13% RFP
45% RFP	45% RFP
62% RFP	62% RFP
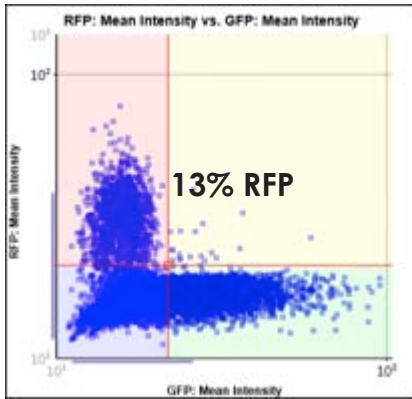
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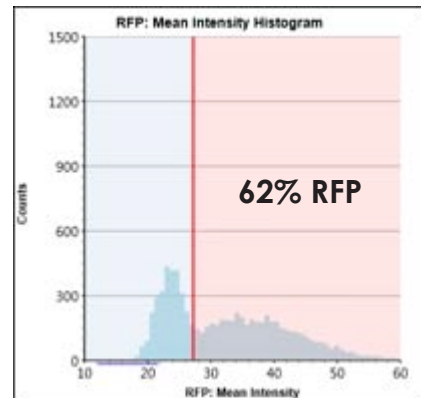
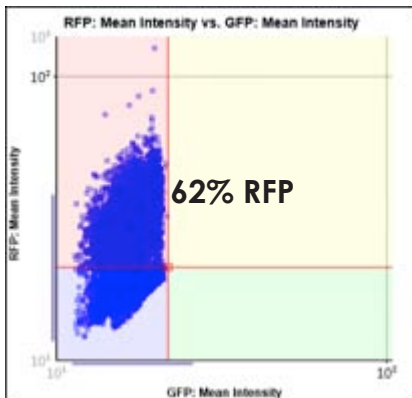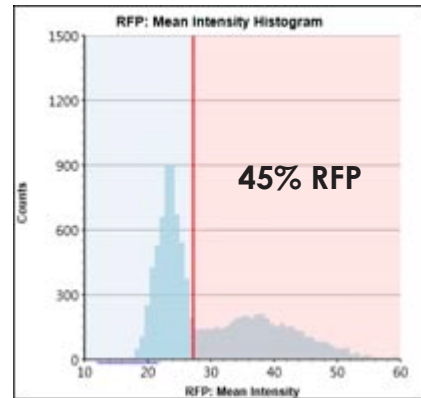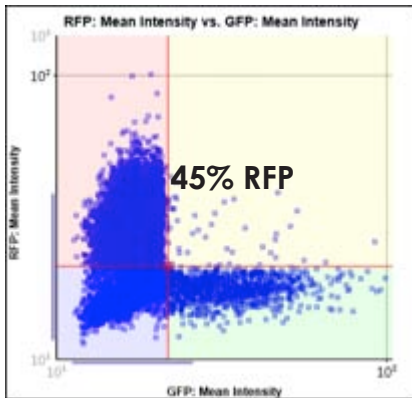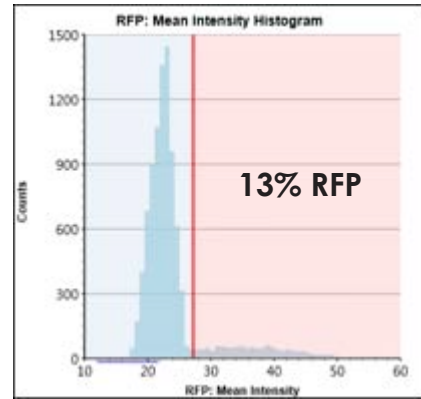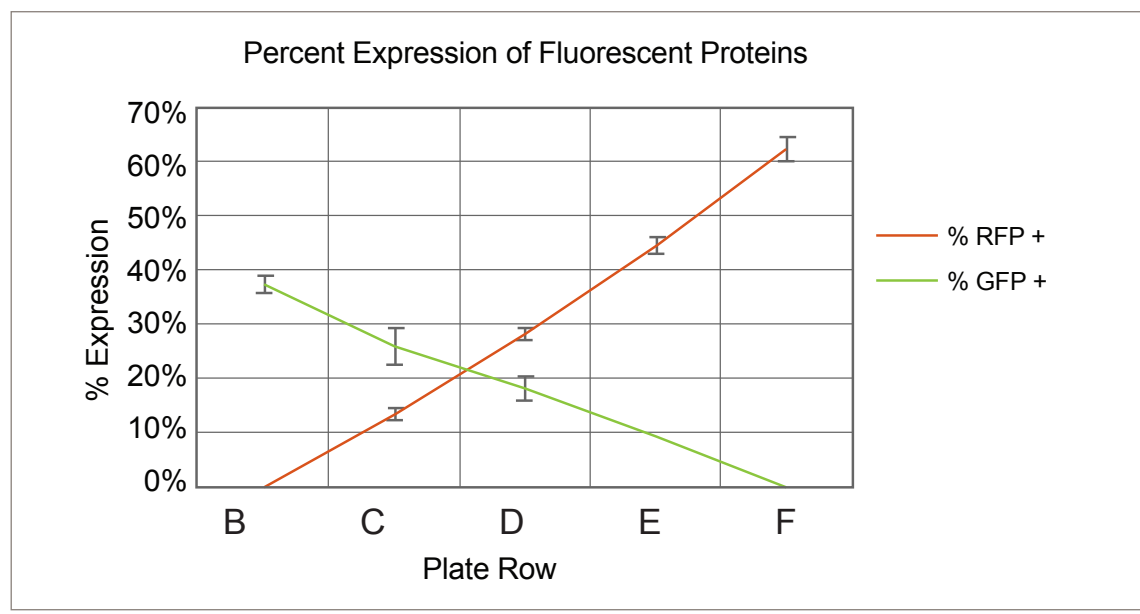## Step 4. Export Results for Each of the Wells in the 96-Well Plate and Produce Data Plot
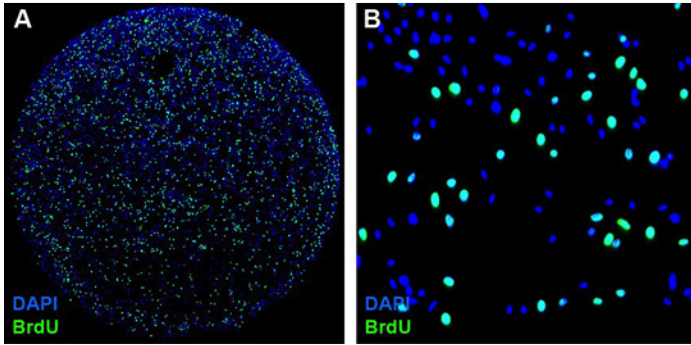
Open Well Level Data in Excel / Calculate AVE & SD / Create Plot with Ave & Std

Measurement Plate Maps

% RFP +	1	2	3	4	5	6	7	8	9	10	11	12	% RFP +	
A													AVE	SD
B		0.2%	0.1%	0.1%	0.1%	0.1%	0.1%	0.1%	0.1%	0.2%	0.1%		0%	0%
C		14.0%	12.9%	13.9%	13.2%	13.5%	13.4%	13.9%	14.2%	13.2%	14.3%		14%	0%
D		28.5%	27.1%	26.1%	28.3%	27.7%	27.7%	28.6%	28.6%	29.3%	30.0%		28%	1%
E		46.4%	42.8%	43.3%	44.5%	44.9%	45.9%	44.2%	43.8%	43.7%	46.0%		45%	1%
F		63.5%	62.3%	62.0%	61.1%	62.0%	60.5%	61.2%	61.4%	61.8%	67.3%		62%	2%
G														
H														

% Hela	1	2	3	4	5	6	7	8	9	10	11	12	% Hela	
A													AVE	SD
B		62.1%	62.7%	63.0%	61.9%	64.4%	60.8%	61.4%	63.0%	61.1%	60.2%		62%	1%
C		61.0%	69.5%	58.6%	60.6%	58.8%	58.0%	58.5%	58.5%	58.4%	55.0%		60%	4%
D		53.8%	53.9%	55.1%	52.6%	54.9%	59.4%	51.6%	51.2%	51.6%	48.2%		53%	3%
E		44.1%	47.3%	46.2%	45.3%	45.2%	43.9%	45.2%	46.4%	46.9%	43.4%		45%	1%
F		36.3%	37.6%	37.8%	38.7%	37.8%	39.2%	38.3%	38.3%	37.7%	32.5%		37%	2%
G														
H														

% GFP +	1	2	3	4	5	6	7	8	9	10	11	12	% GFP +	
A													AVE	SD
B		37.4%	37.1%	36.7%	37.8%	35.3%	38.9%	38.3%	36.6%	38.5%	39.5%		38%	1%
C		24.7%	17.3%	26.9%	25.9%	27.1%	28.1%	27.2%	26.9%	28.0%	29.8%		26%	3%
D		17.2%	18.3%	18.4%	18.6%	17.0%	12.5%	19.1%	19.6%	18.5%	21.0%		18%	2%
E		9.0%	9.4%	10.0%	9.7%	9.4%	9.6%	9.9%	9.3%	8.9%	9.5%		9%	0%
F		0.0%	0.0%	0.0%	0.0%	0.0%	0.0%	0.1%	0.0%	0.2%	0.0%		0%	0%

Percent Expression of Fluorescent Proteins
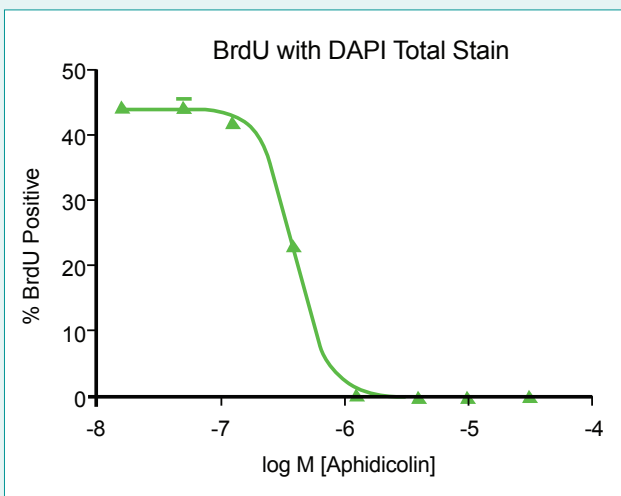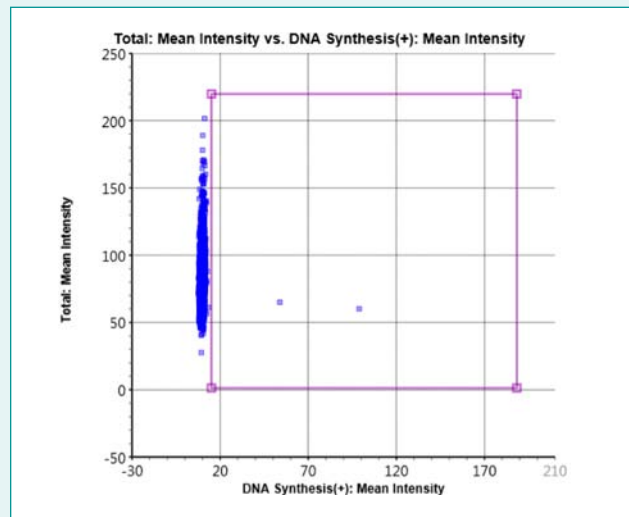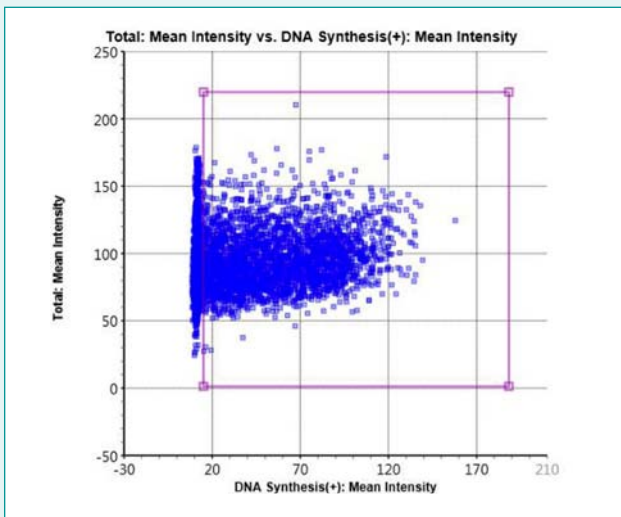
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Cell Proliferation Assay Using BrdU Incorporation

Image Data

Gated Data

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Cell Cycle Analysis of Adherence Cells in 96-Well Plates Using Celigo

Cells were stained with DAPI, a marker for DNA synthesis. The multi-fluorescent channel analysis and gating interface allows identification of cells in the G0/G1, S-Phases and G2/M phases of the cell cycle.

Cell Cycle - Method 1: Measure DNA Content Using DAPI

|	Control	150 nM Nocodazole	50 µM Nocodazole
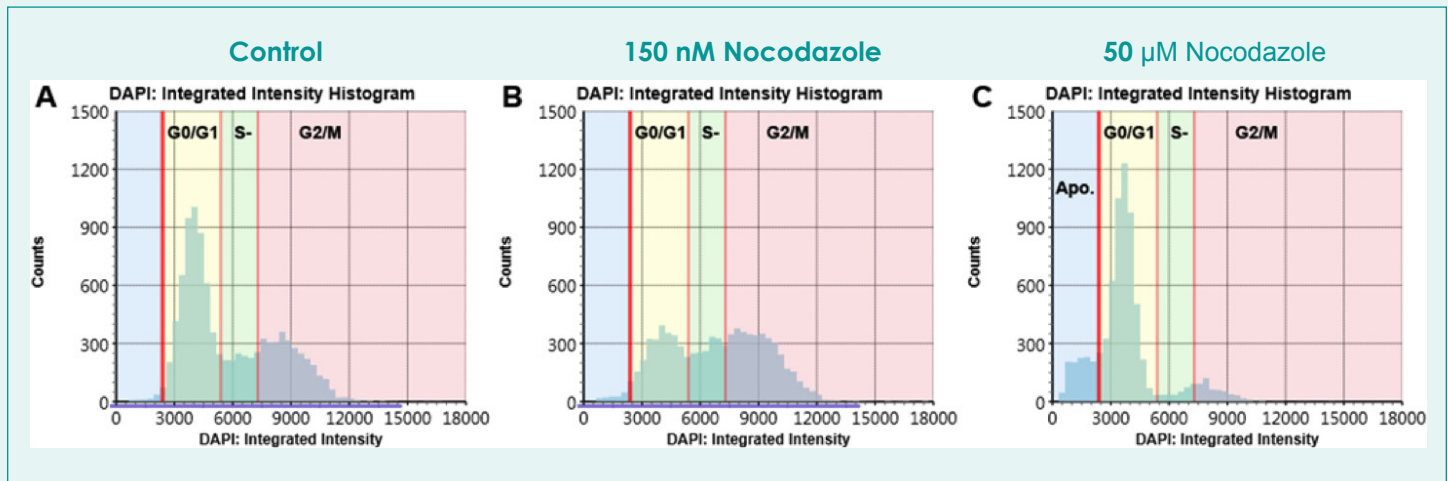
Cell Cycle-Method 2: Using DAPI and BrdU

- Cell cycle analysis is examined by measuring the amount of DNA per cell
- Incorporation of DAPI and BrDU
- Double peak characteristic peak of cell cycle consisting of G0/G1, S and G2/M phases

	Control	100 µM Aphidicolin
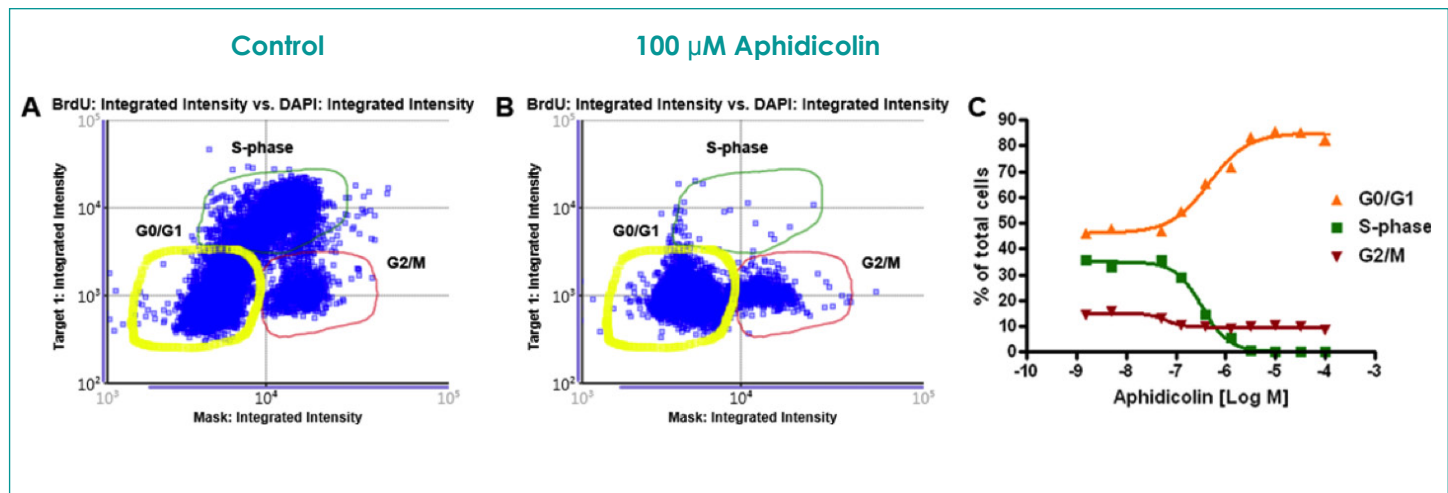
Imaging data can be used to generate dose response curves to characterize drug activities on the cell cycle (right panel).

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Cell Viability Determined by Staining Cells with Live and Dead Cell-Specific Dyes

Briefly, cells are simultaneously stained with a mixture of calcein AM, propidium iodide, and Hoechst 33342 for respective staining of live, dead and all cells. Images are acquired and analyzed using the Celigo software. Markers are identified in each fluorescent channel and for each well of a microtiter plate, live and dead cell counts as well as the percentage of live and dead cells are automatically reported.
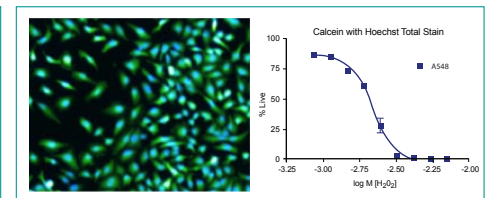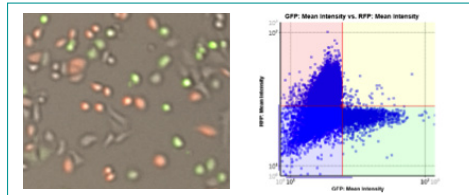
Cell Counting in 96-Well Plate

Green: Live, Dead: Red

Green: Live, Blue: Total
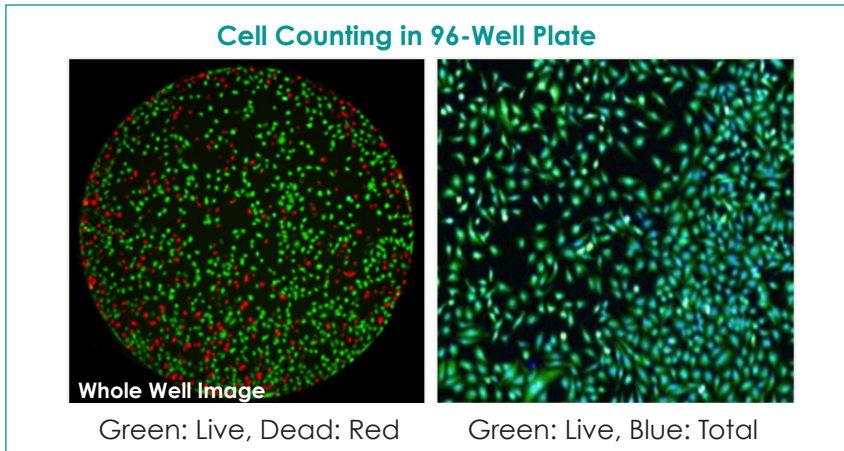
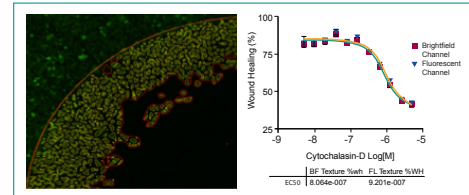## Cell Cycle by BrdU
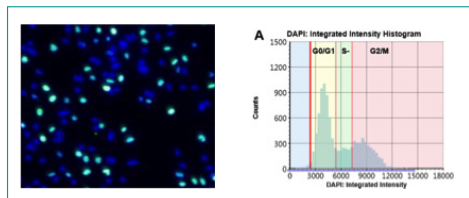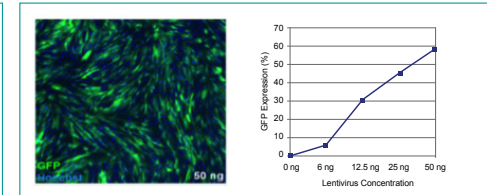

Apoptosis - PS Externalization
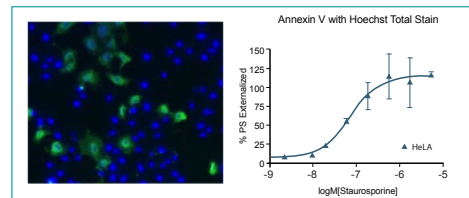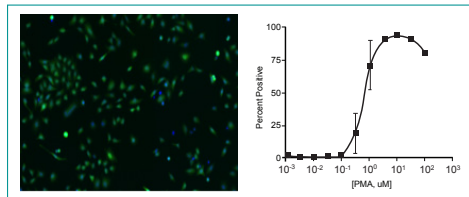

Cell Viability

Transfection Efficiency
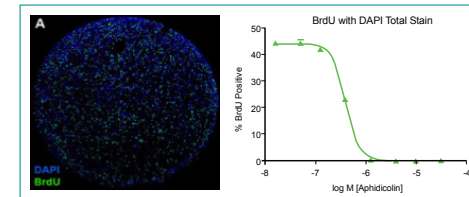

Migration

Viral Infection

ERK Phosphorylation

DNA Synthesis

- **Receptor internalization**
- **Phagocyosis**
- **Receptor detection: CD71, CD54 ICMA-1**
- **ERK Phophorylation**
- **Nuclear antigen detection**

Cellometer®
Simply Counted

Celígo S
Image Cytometer

Nexcelom Bioscience

For more information, visit
www.nexcelom.com

Contact us at:
Nexcelom Bioscience
360 Merrimack Street, Building 9
Lawrence, MA 01843, USA

Email: info@nexcelom.com
Phone: 978.327.5340
Fax: 978.327.5341

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